

# Flexibility Prediction: Preserving Detailed Balance in Side Chain and Backbone Movers

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RosettaCON - Wednesday, August 4, 2010

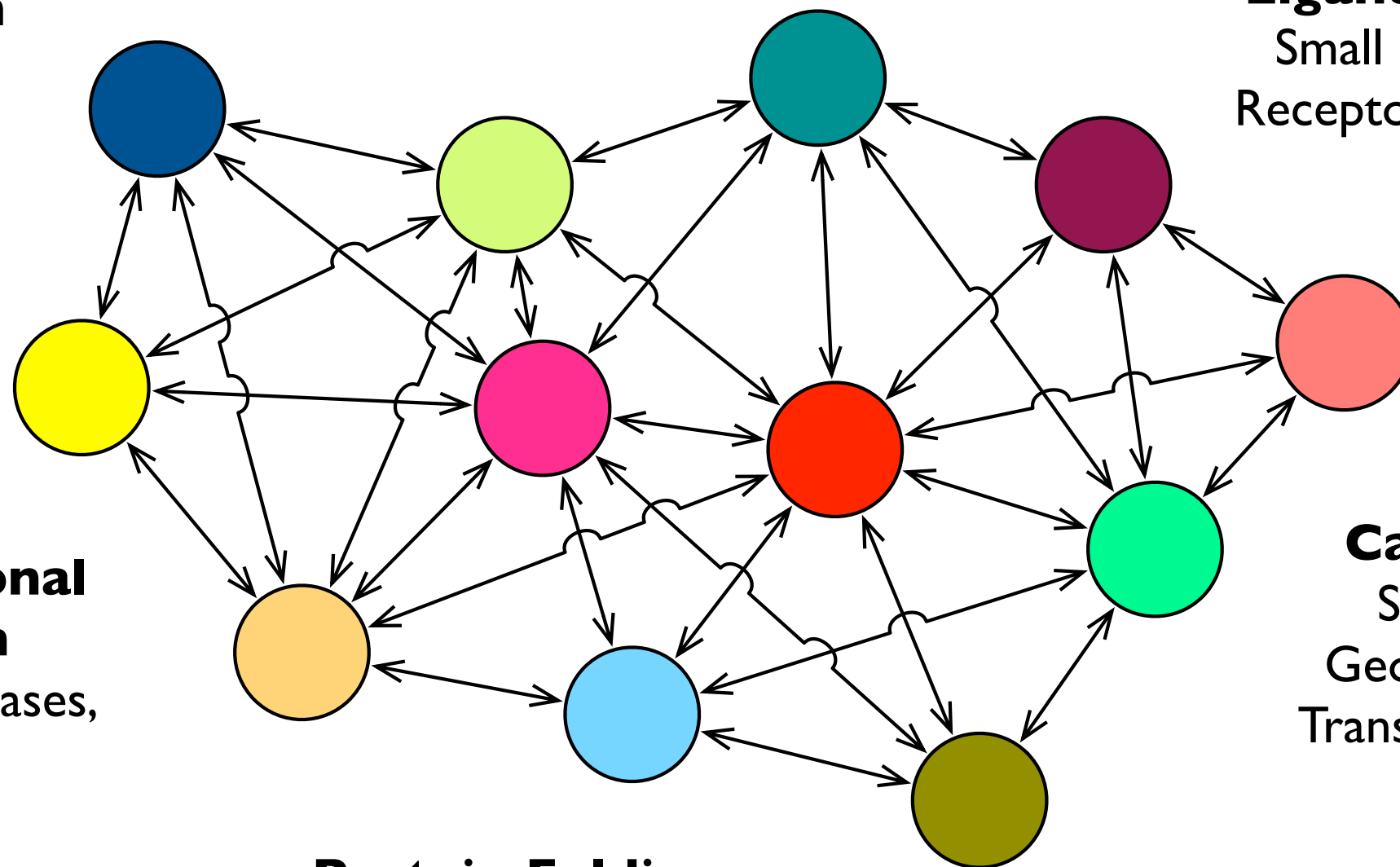
# Predicting Flexibility With Monte Carlo Simulations

## Protein Protein Interaction

Free Energy  
Cost of Binding

## Ligand Binding

Small Molecule/  
Receptor Flexibility



## Post Translational Modification

Accessibility to Kinases,  
Proteases, etc.

## Catalysis

Stabilize  
Geometry of  
Transition State

**Protein Folding**  
Free Energy vs.  
Potential Energy Basin

# Rigorous Monte Carlo Must Preserve Detailed Balance

Analogous to the Law of Mass Action:



Population

Transition Probability

$$K_{eq} = \frac{[B]}{[A]} = \frac{k_{forward}}{k_{reverse}}$$

Solution #2:

$$a_{AB} = \frac{1}{q_{AB}} e^{(E_A - E_B)/kT}$$

$$p_{AB} = q_{AB} a_{AB}$$

$$\frac{\pi_B}{\pi_A} = \frac{p_{AB}}{p_{BA}} = \frac{q_{AB} a_{AB}}{q_{BA} a_{BA}}$$

$$\frac{\pi_B}{\pi_A} = \frac{p_{AB}}{p_{BA}} = \frac{q_{AB} a_{AB}}{q_{BA} a_{BA}}$$

Mover Proposal Density

Metropolis Criterion

Problem:  $q_{AB} \neq q_{BA}$     Solution #1:  $q_{AB} = q_{BA}$

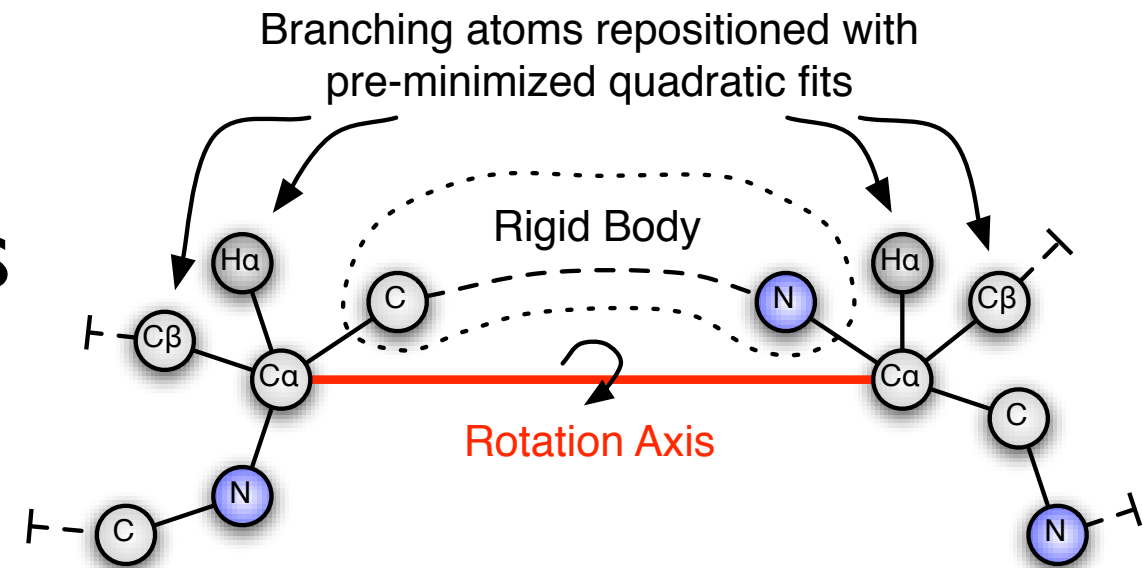
# Movers Using Solution #1

- BackrubMover

- Problem: Branching atoms sampled nonuniformly

- Solution:

Turn off branching atom sampling

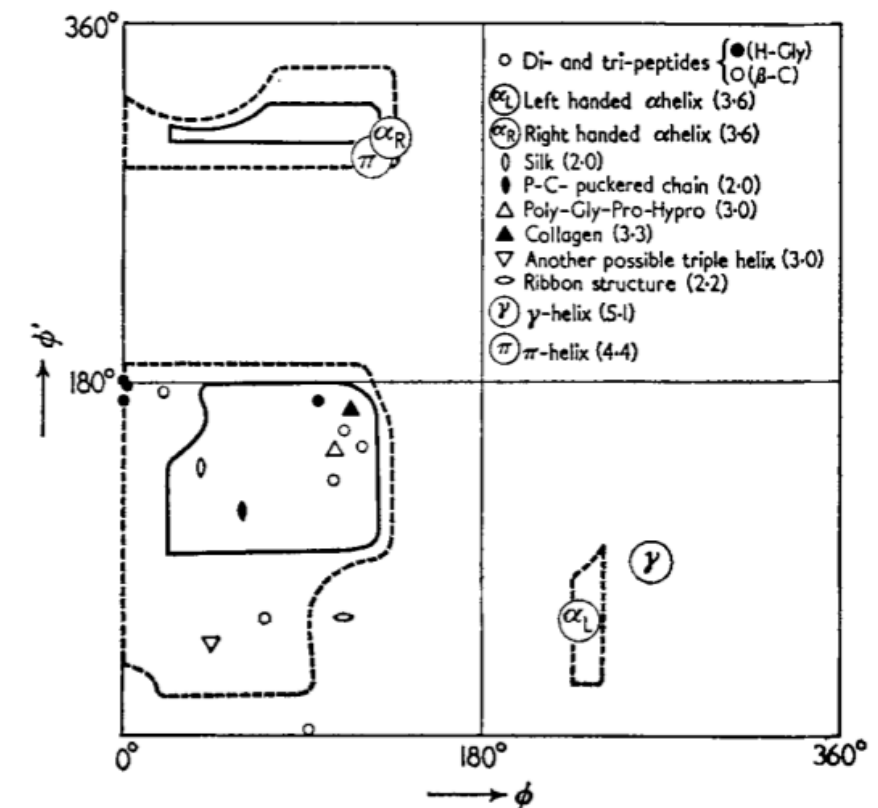


- SmallMover and ShearMover

- Problem: Phi/Psi angles biased by Ramachandran plot

- Solution:

Turn off Ramachandran biasing



# Rotamer Packing vs. SidechainMover

## Rotamer Packing

high probability  
rotamer wells  
sampled uniformly

discrete chi angles  
sampled within  
rotamer wells

always selects  
a random rotamer  
configuration

you have to worry  
about library resolution  
(-ex1, -ex2, etc.)

## SidechainMover

all rotamer wells  
sampled with  
PDB probability

chi angles sampled  
continuously from a  
Gaussian distribution

can also sample within  
current rotamer well or  
all chi angles uniformly

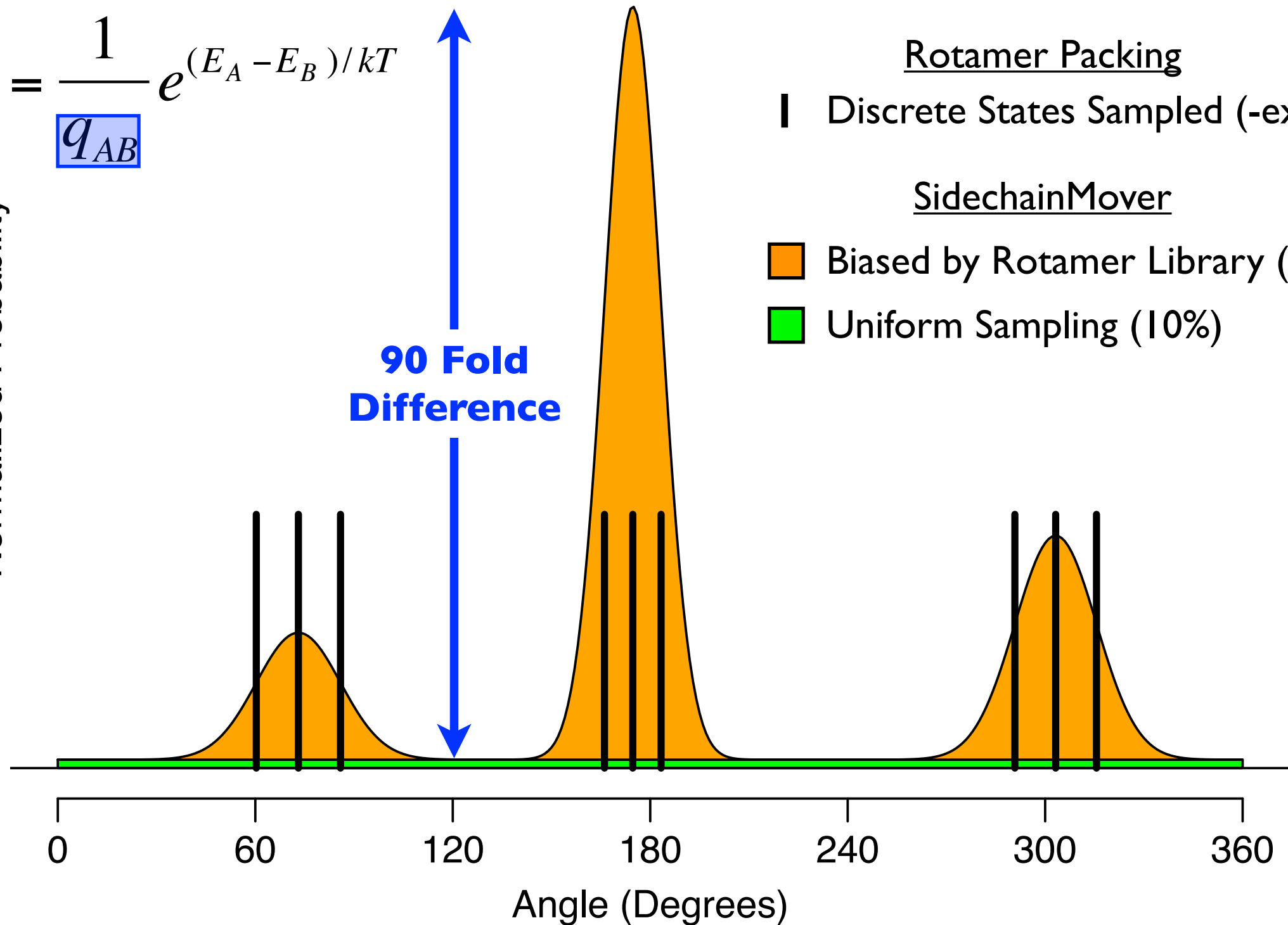
all possible chi angle  
combinations  
are sampled

# Rotamer Packing vs. SidechainMover

Solution #2: Valine chi 1 Sampling ( $\phi=-80$ ,  $\psi=-30$ )

$$a_{AB} = \frac{1}{q_{AB}} e^{(E_A - E_B)/kT}$$

Normalized Probability



# Testing Detailed Balance

- Strategy: Run simulations and make sure movers generate expected populations.
- Easiest population to predict: Empty scoring function:  
$$a_{AB} = \frac{1}{q_{AB}} e^{(E_A - E_B)/kT}$$
- Use simplified systems for nightly testing:
  - SidechainMover: Single residue poses
  - Other movers: 8 residue polyalanine

# SidechainMover Example: Arginine Angle Histograms

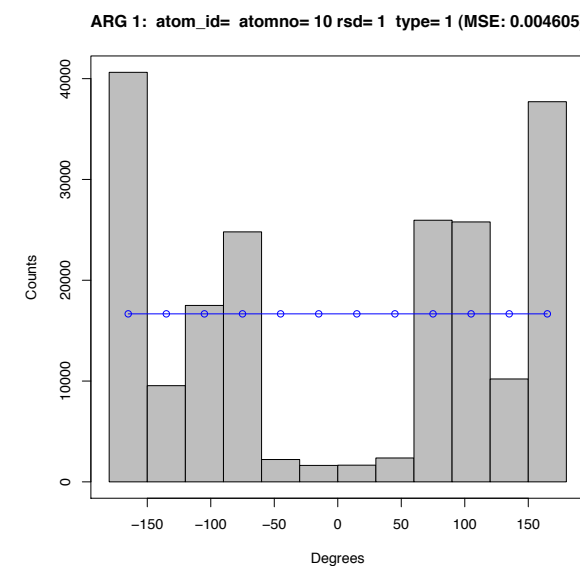
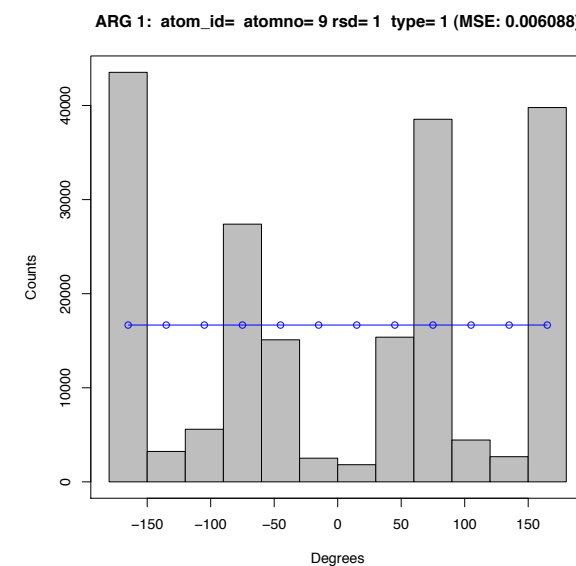
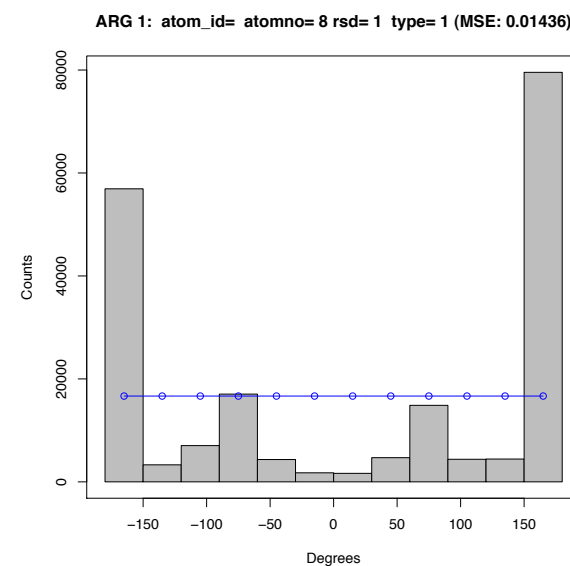
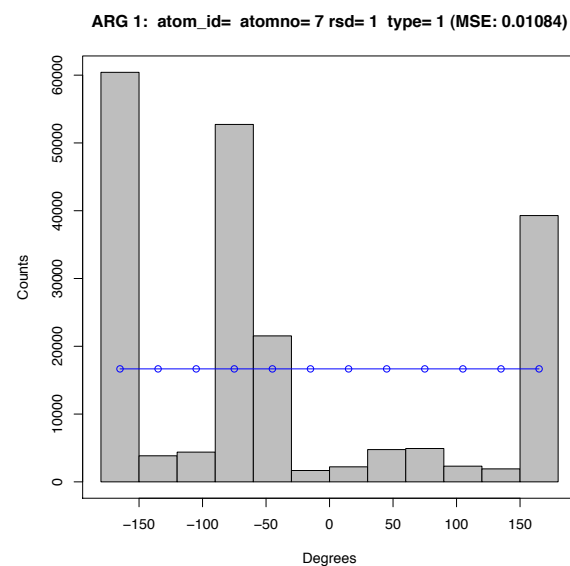
chi 1

chi 2

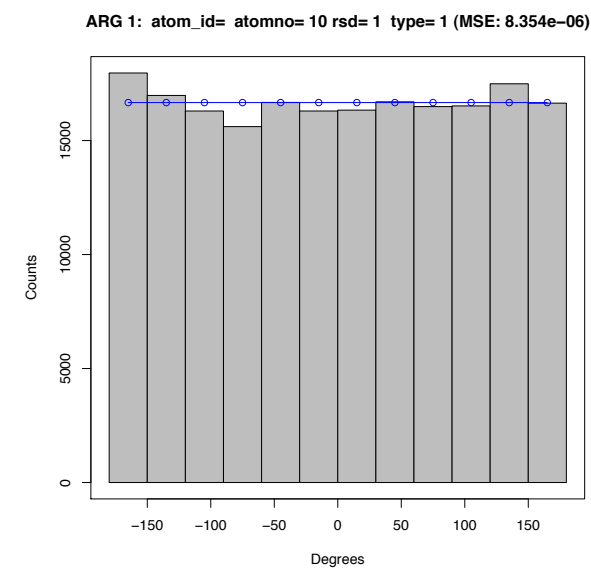
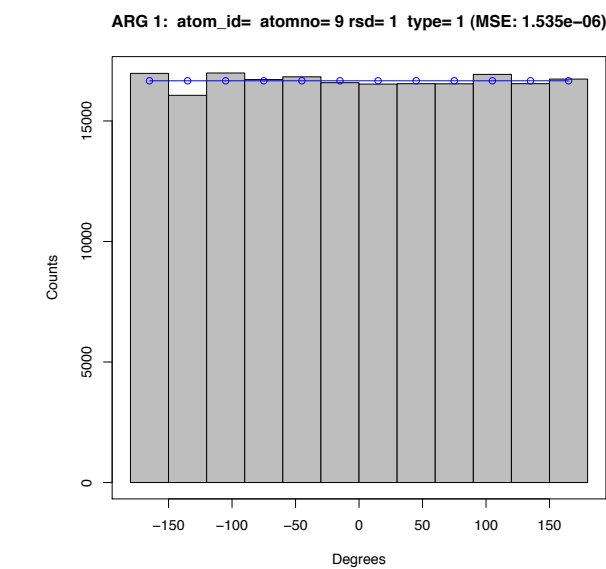
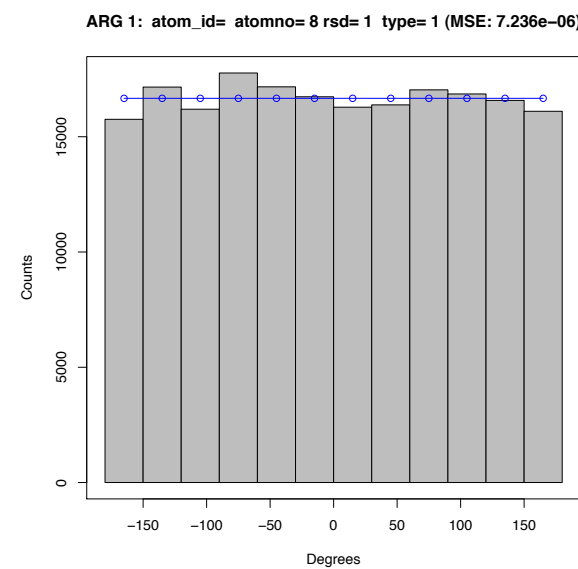
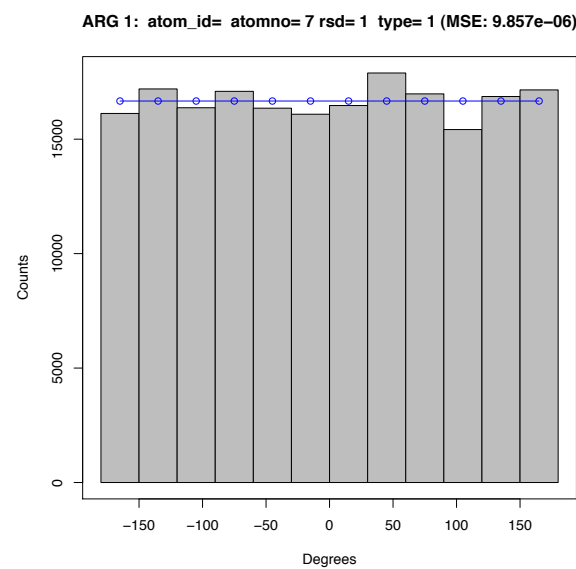
chi 3

chi 4

Without  
Detailed Balance

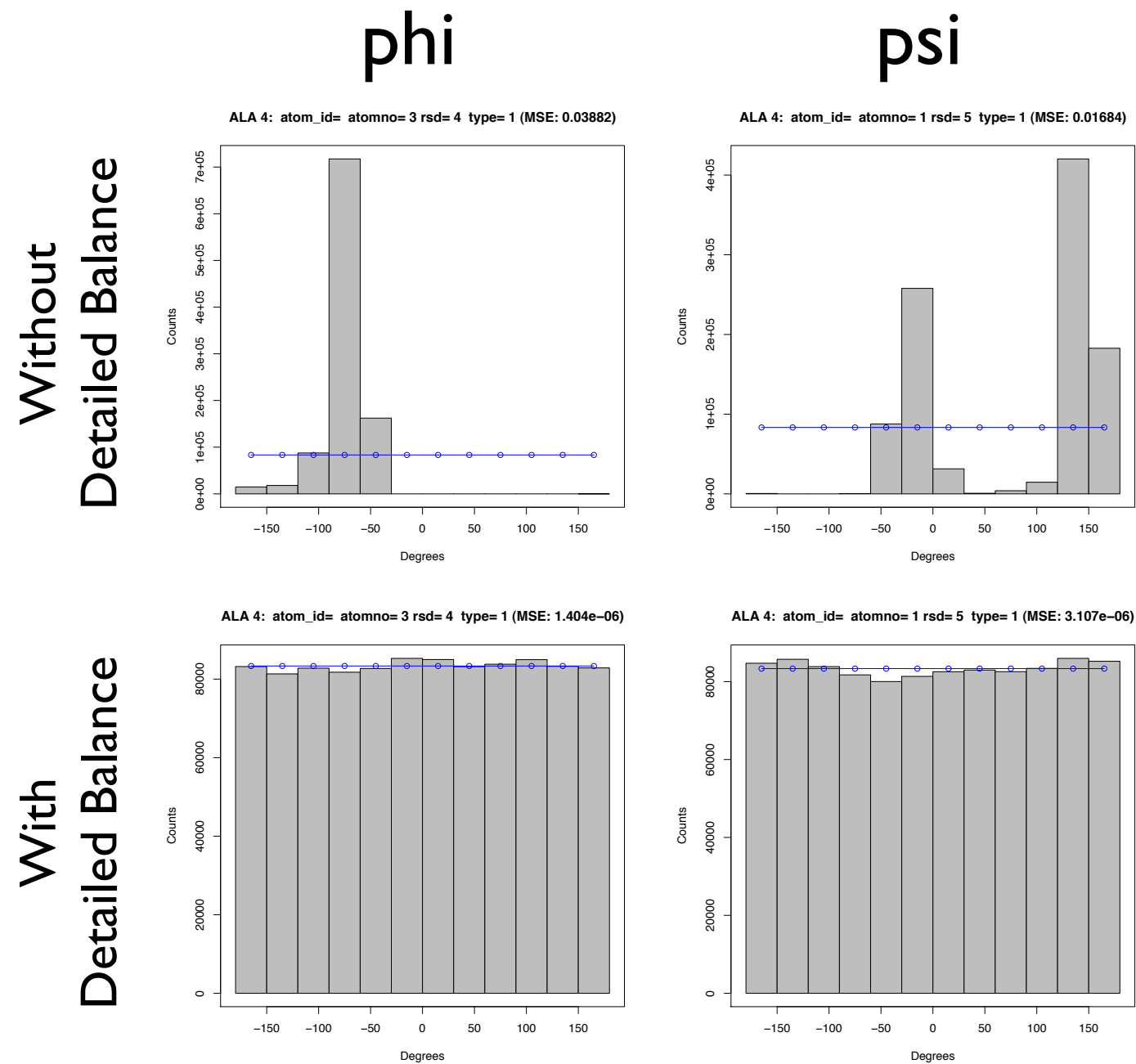


With  
Detailed Balance





# SmallMover Example: Polyalanine Residue 4



# BackrubMover Example: Polyalanine Residue 4

phi

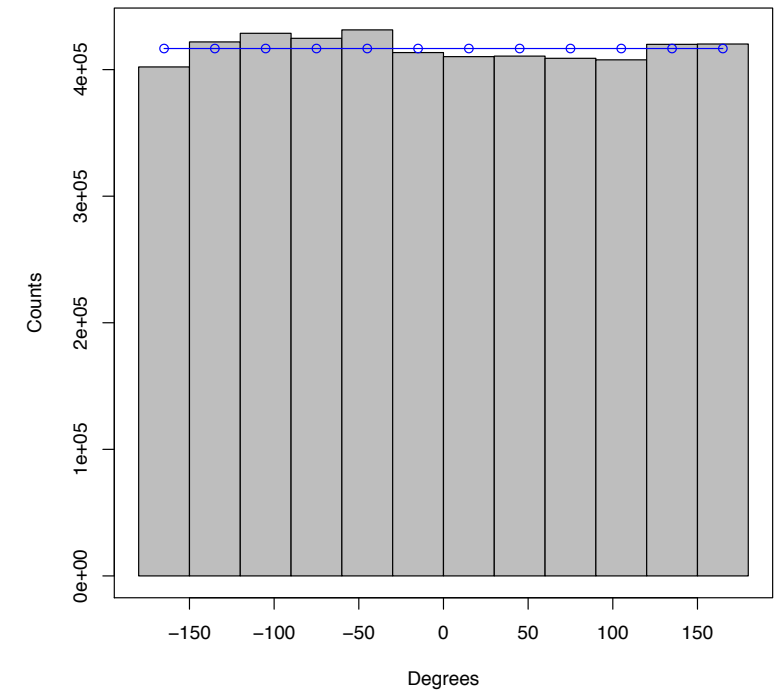
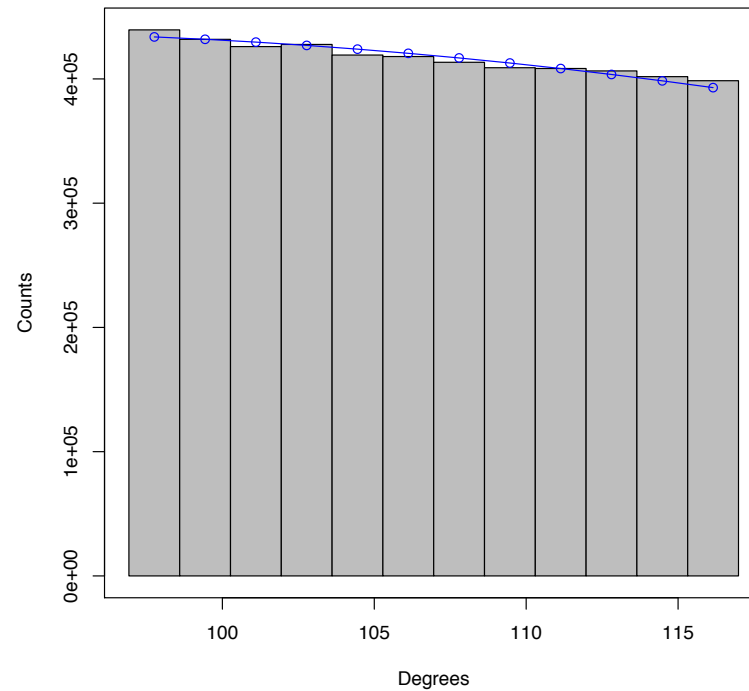
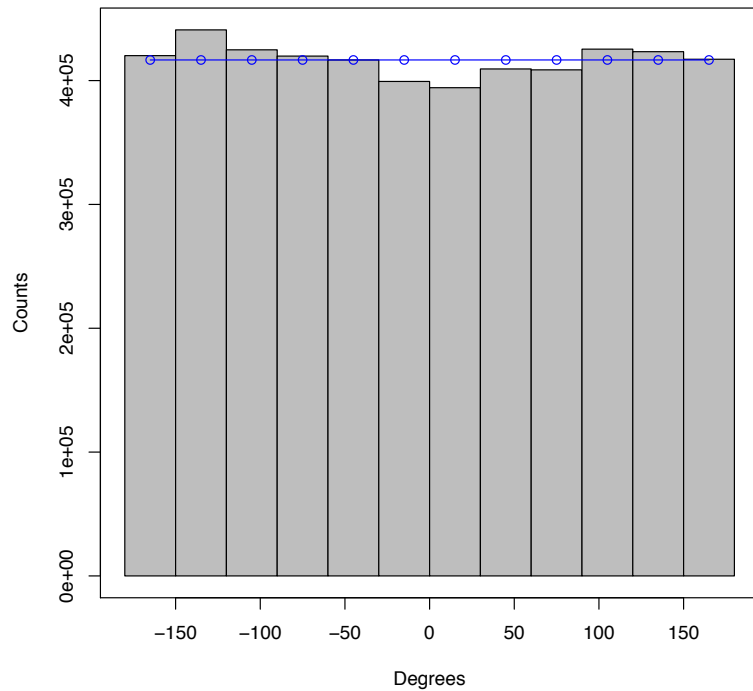
N-CA-C bond angle

psi

ALA 4: atom\_id= atomno= 3 rsd= 4 type= 1 (MSE: 5.731e-06)

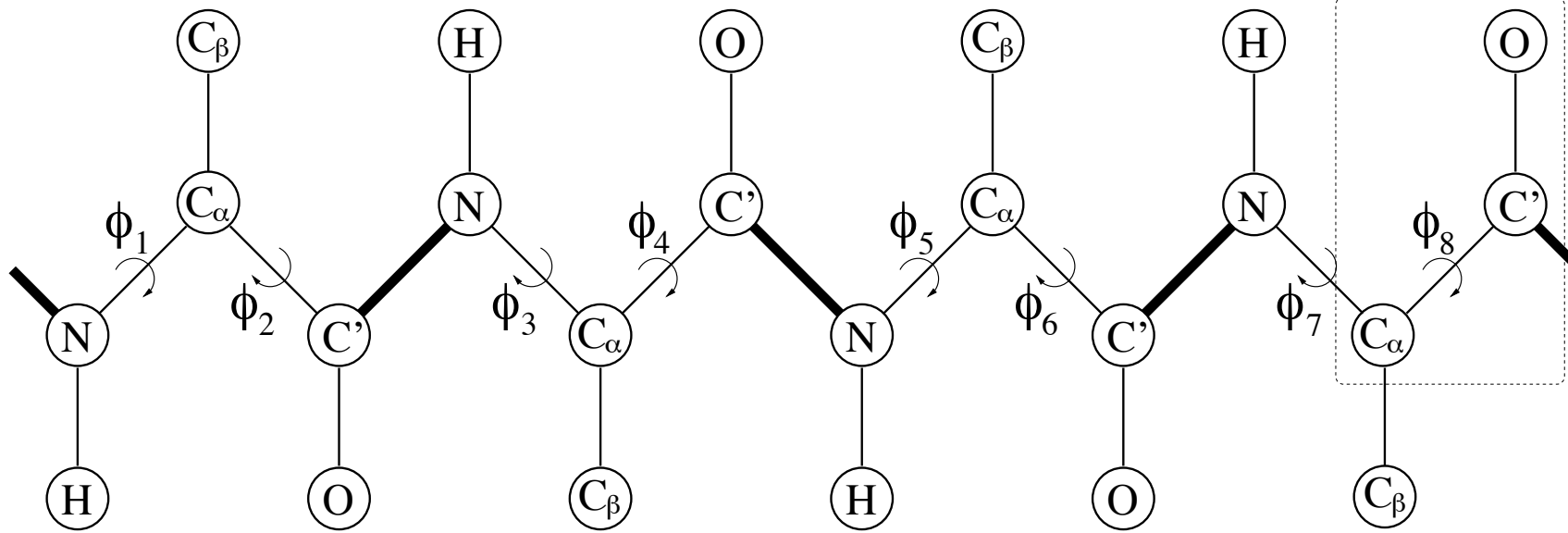
ALA 4: atom\_id= atomno= 3 rsd= 4 type= 2 (MSE: 5.06e-07)

ALA 4: atom\_id= atomno= 1 rsd= 5 type= 1 (MSE: 3.053e-06)



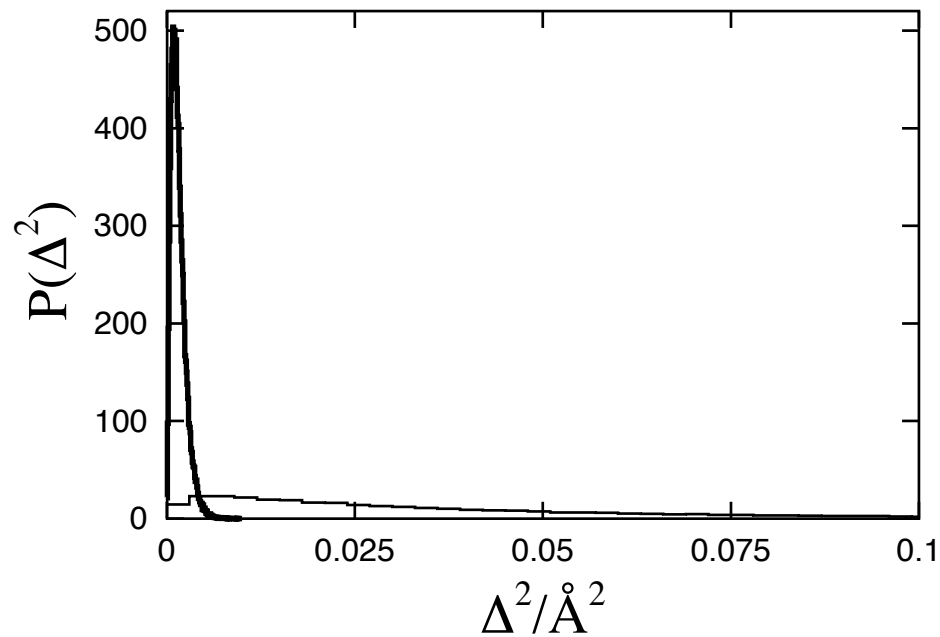
With or Without  
Detailed Balance

# Yuan Liu: Backbone Biased Gaussian Mover

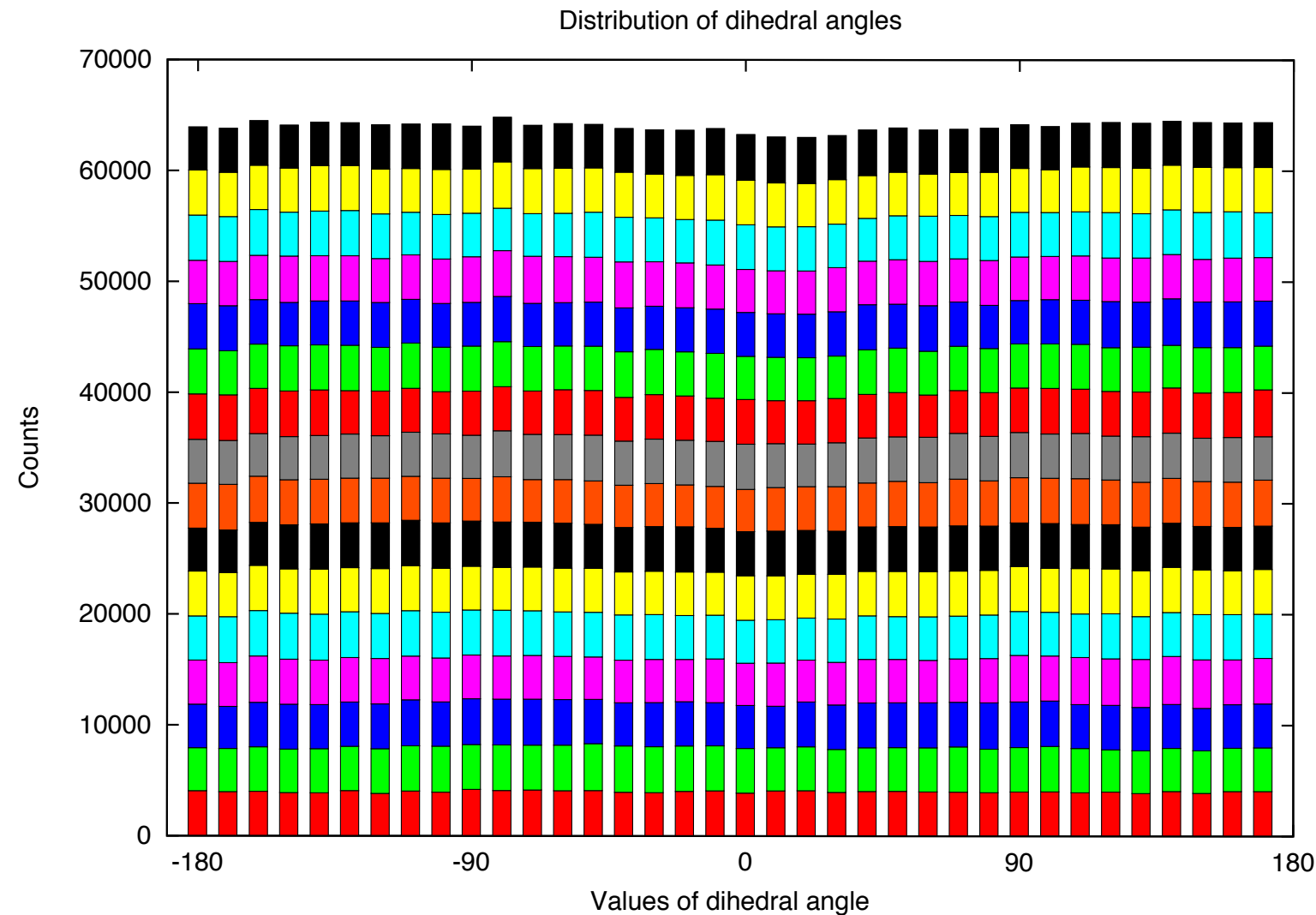


$$\Delta^2 = \sum_{I=1}^3 (\delta \vec{r}_I)^2$$

C<sub>α</sub>, C' and O are the first three atoms of downstream rigid body



Distribution of displacement of the downstream(thick line) is much more local than distribution of randomly perturbed dihedral angles(thin line)



# What kT to Approximate Room Temperature Flexibility?

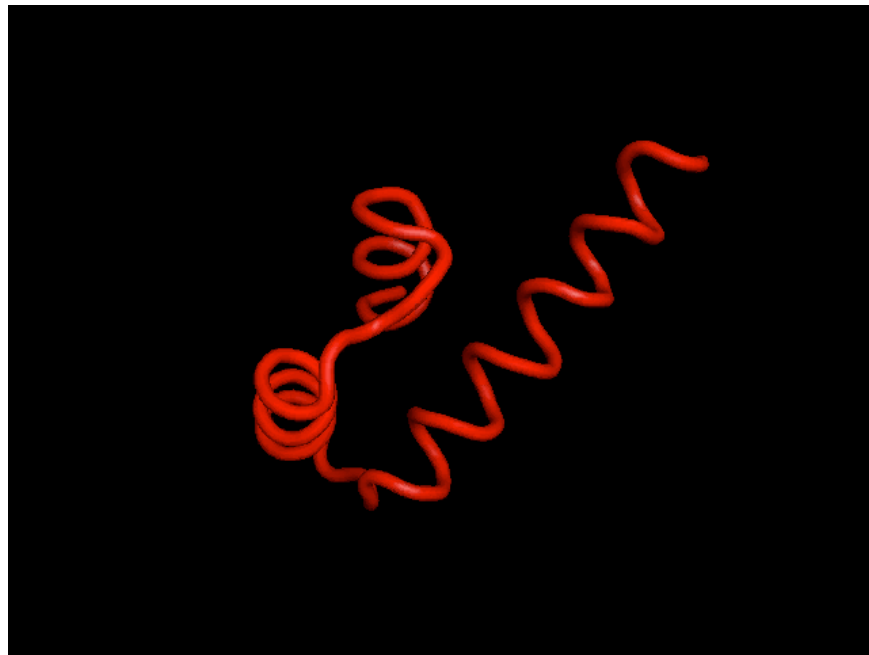
$$a_{AB} = \frac{1}{q_{AB}} e^{(E_A - E_B)/kT} ?$$

- Set an upper bound on Rosetta “room temperature” by running extended simulations of proteins
- Pick the temperature where none unfold

# All 6 Proteins Unfold at 0.6 kT

**kT = 0.6**

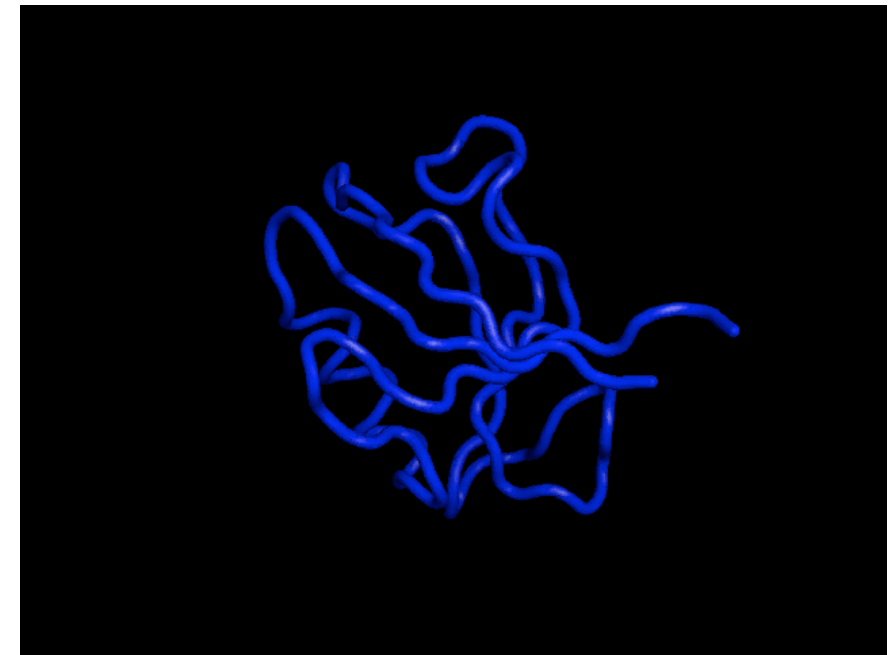
**1B72A**



**1CSP**



**1MFG**



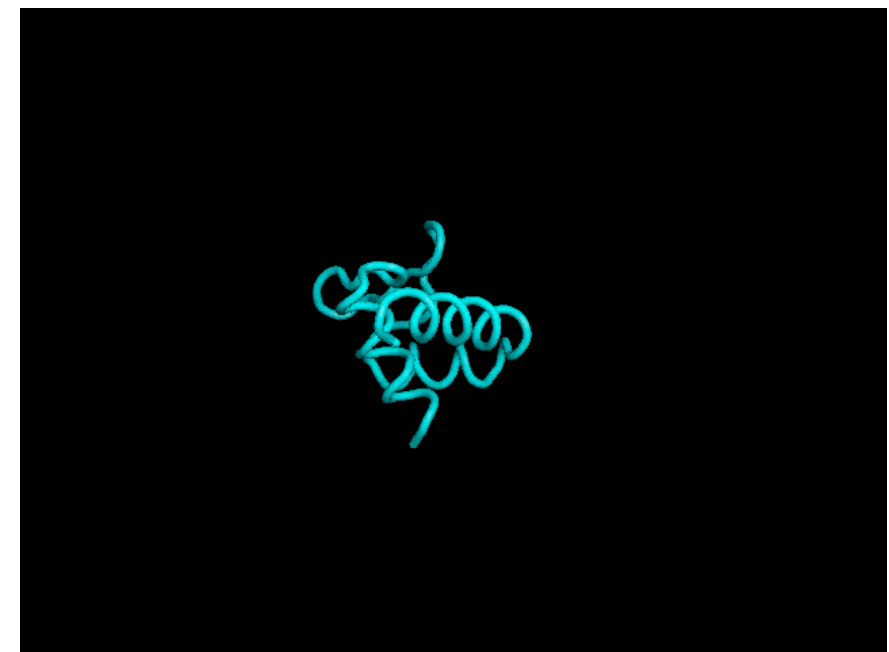
**1SHFA**



**1TIG**



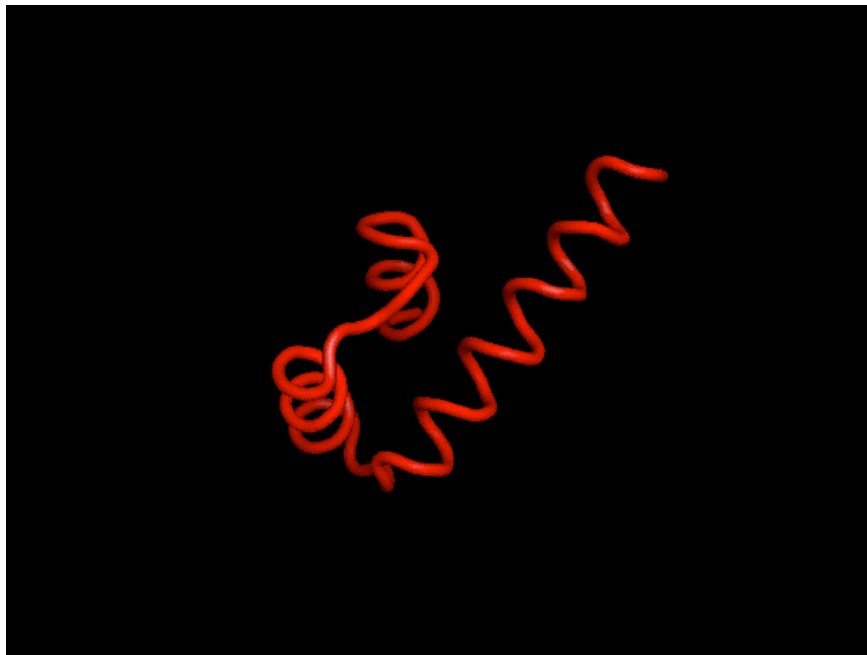
**2REB2**



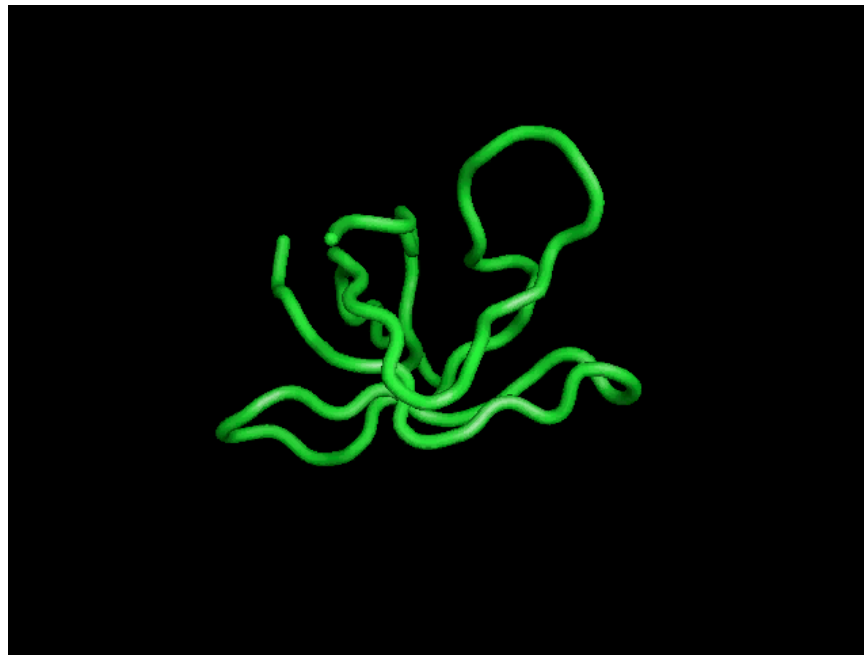
# All 6 Proteins Are Stable at 0.3 kT

**kT = 0.3**

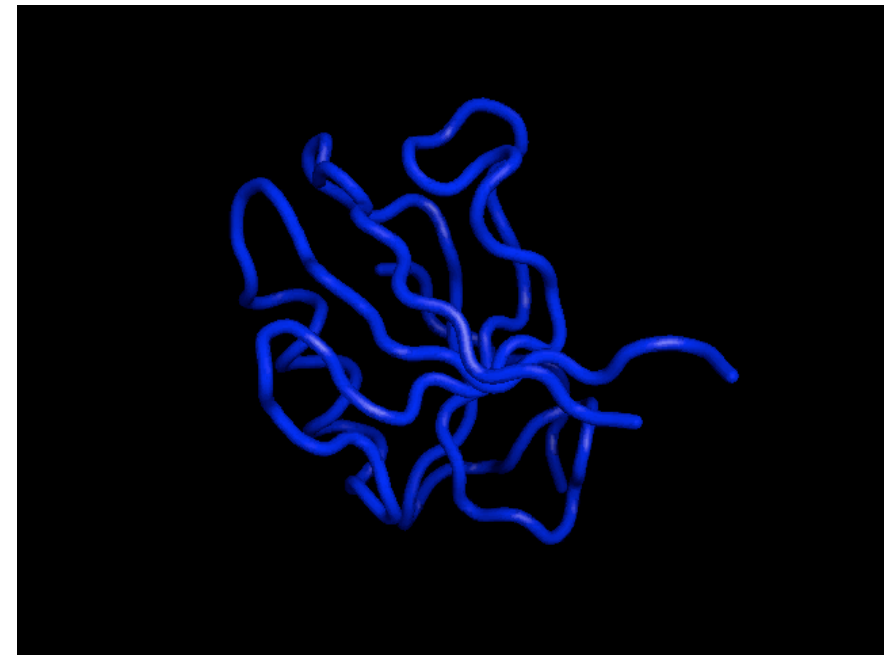
**1B72A**



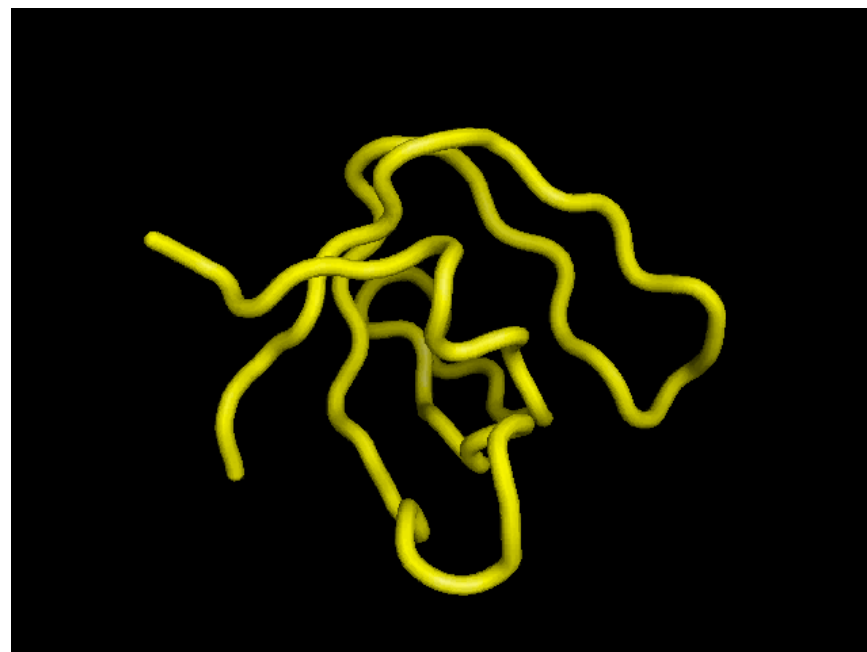
**1CSP**



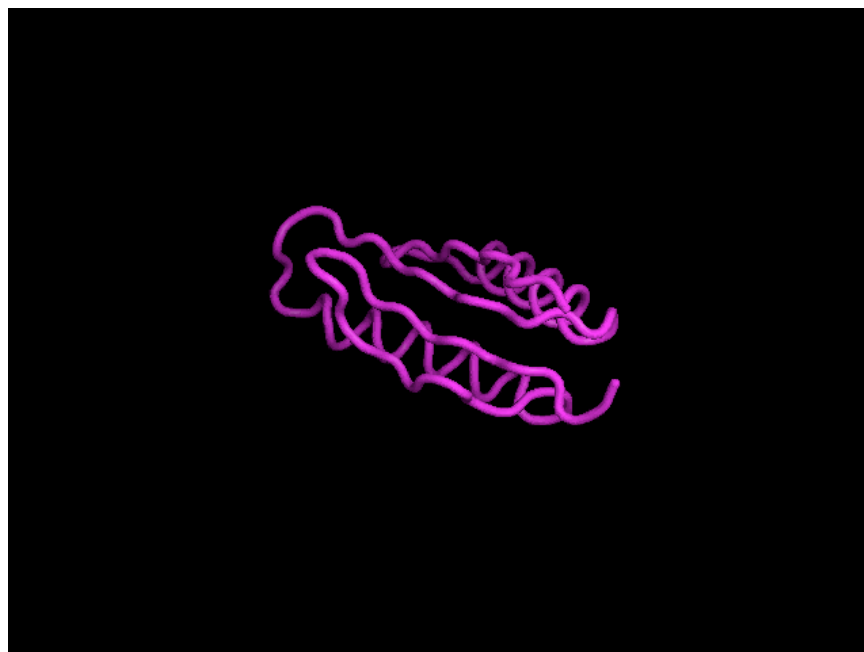
**1MFG**



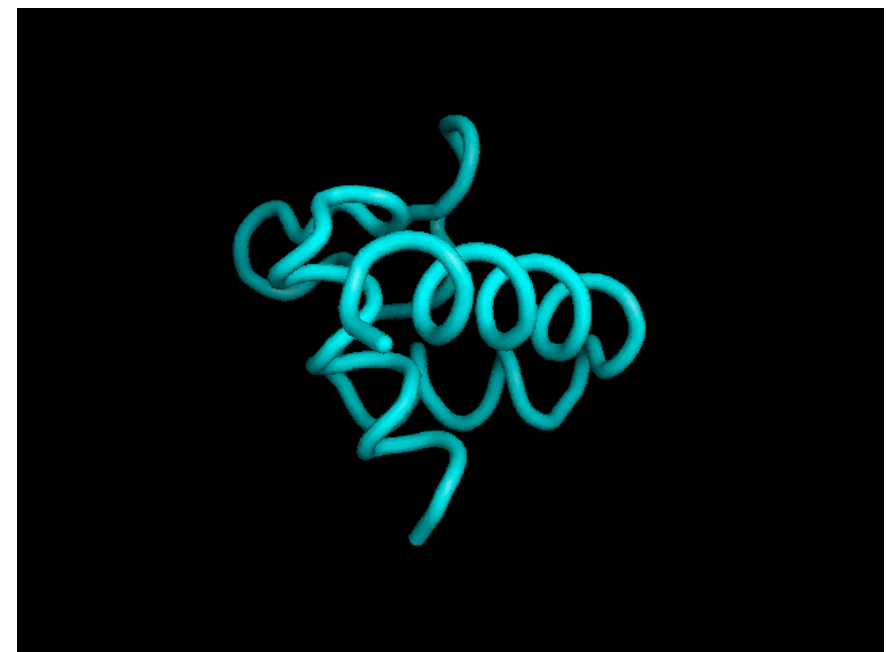
**1SHFA**



**1TIG**



**2REB2**



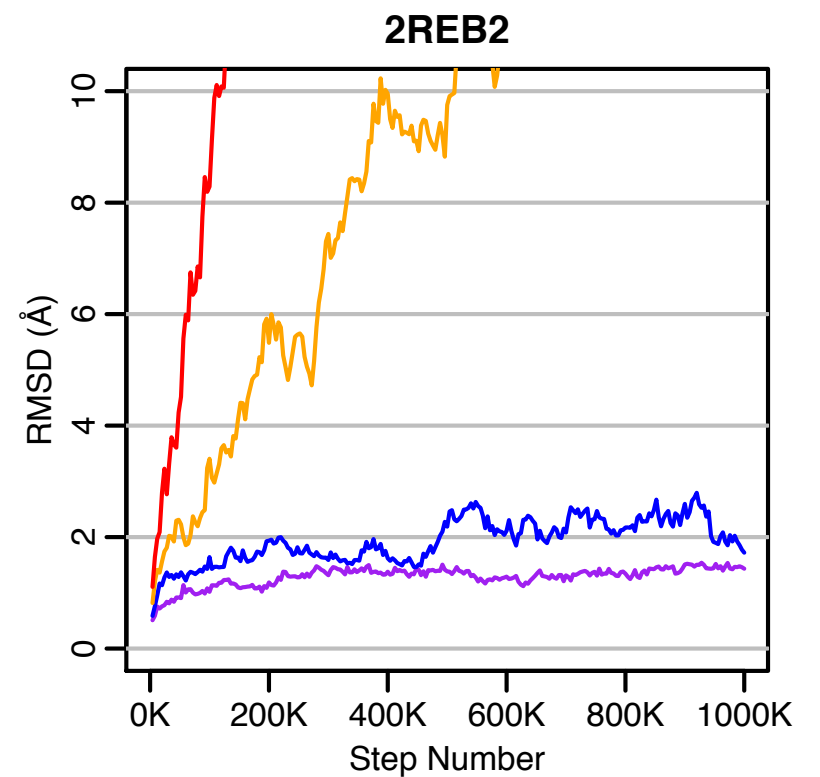
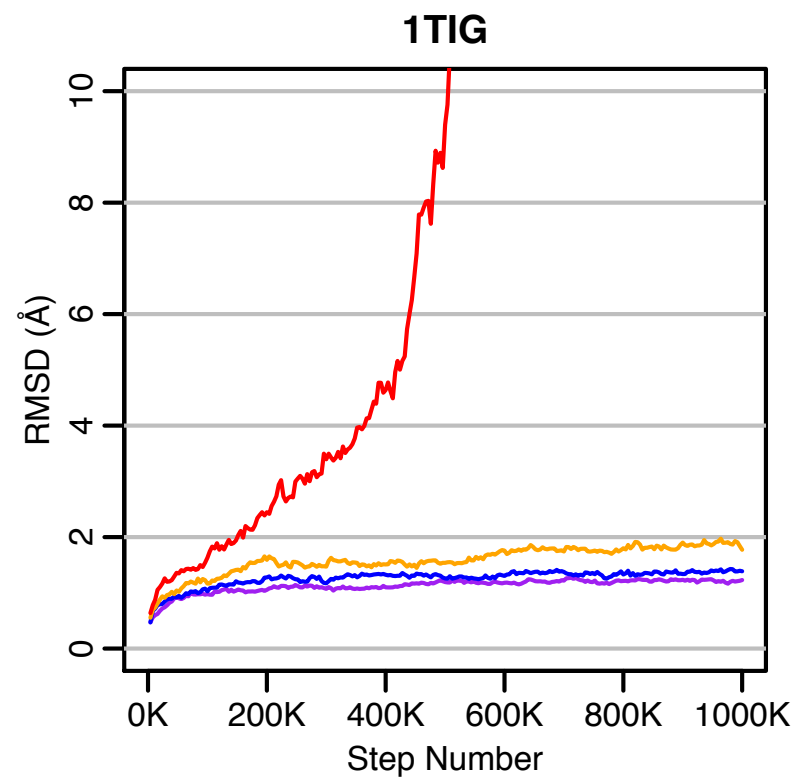
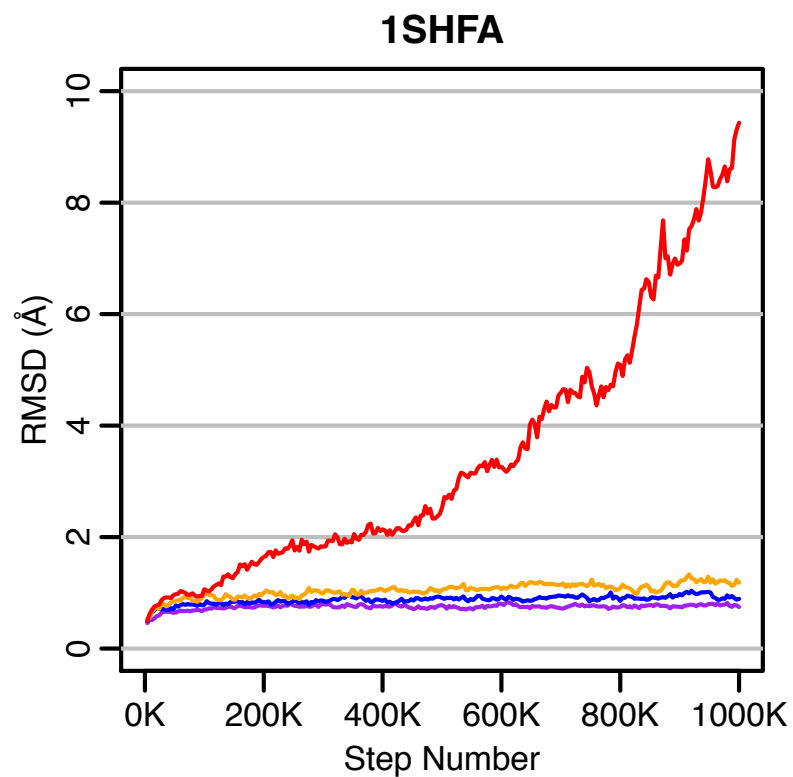
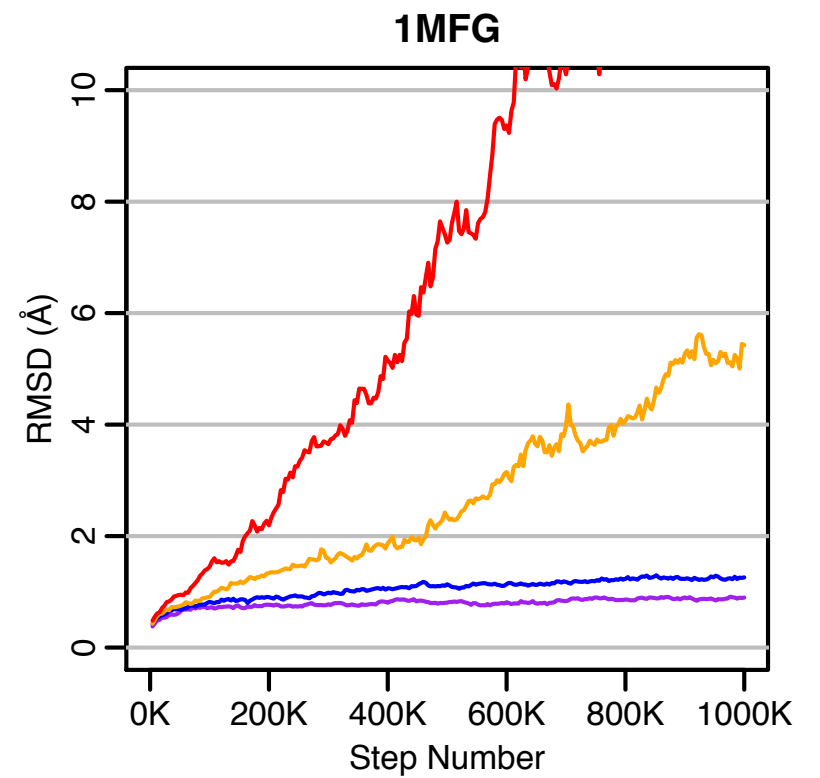
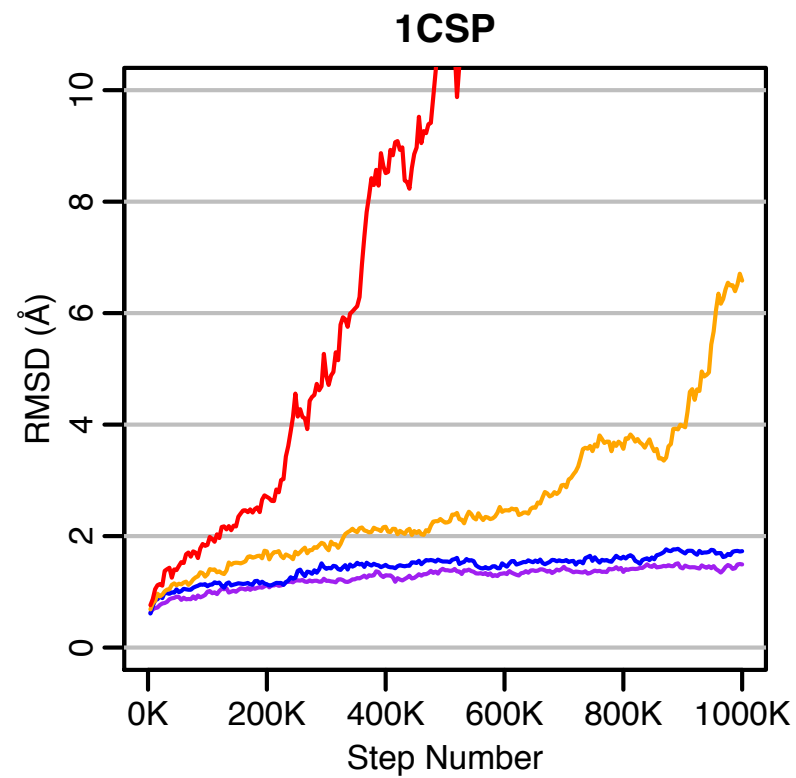
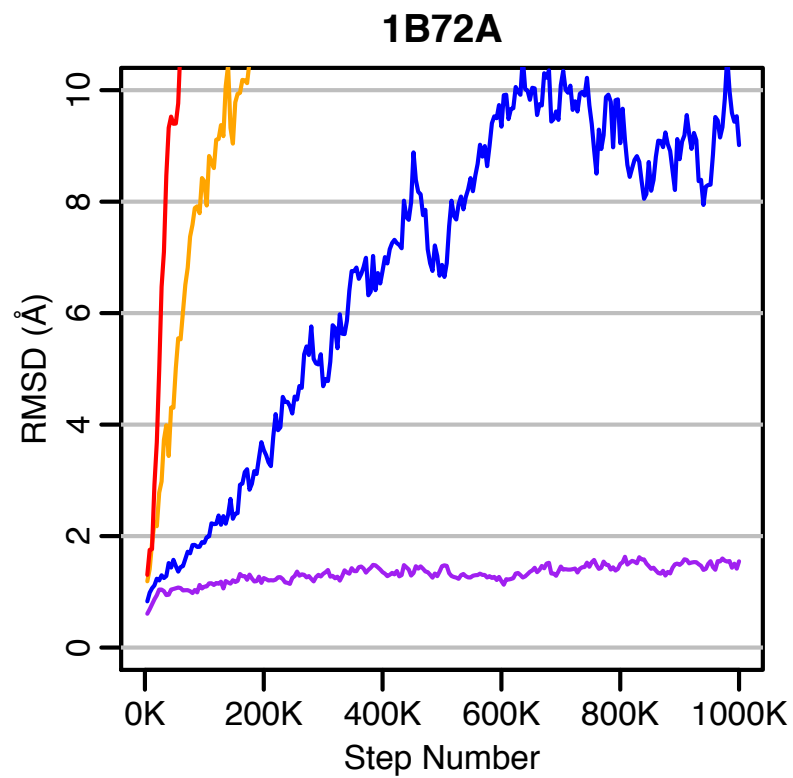
# I/O Simulation Mean RMSD

**kT = 0.3**

**kT = 0.4**

**kT = 0.5**

**kT = 0.6**



# Model System: Cyclophilin A

- Catalyzes proline cis-trans isomerization
- Localized side chain/backbone flexibility near the active site has been observed crystallographically and shown to be temperature dependent
- Studies have shown correlations between the rate of protein motion and catalysis
- Even if there is no causation, this is still an interesting system because of the crystallographic motion

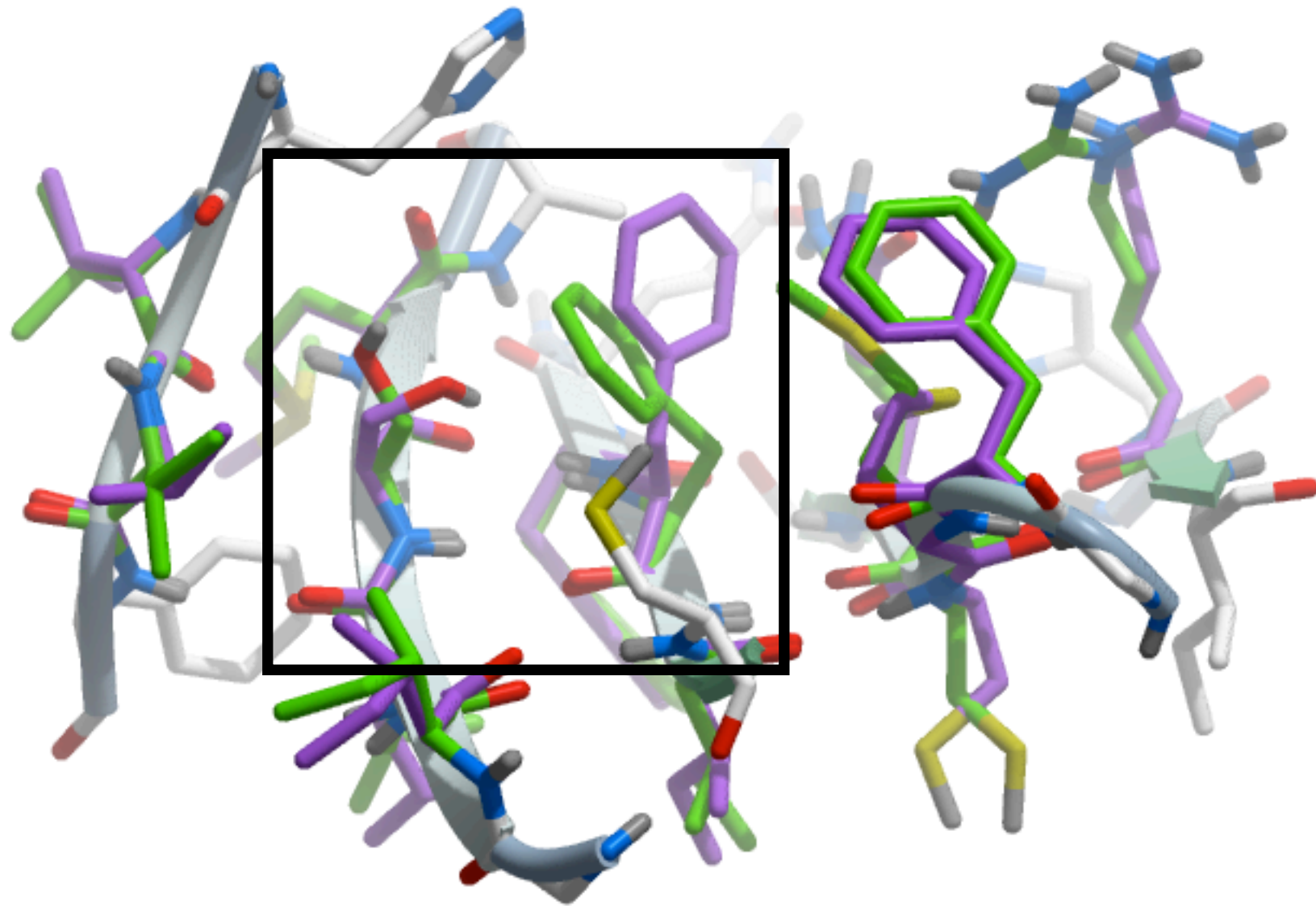


# Crystallography shows alternate conformations at room temperature



Most of the alternate conformations occur in the 2 beta sheets of the beta sandwich

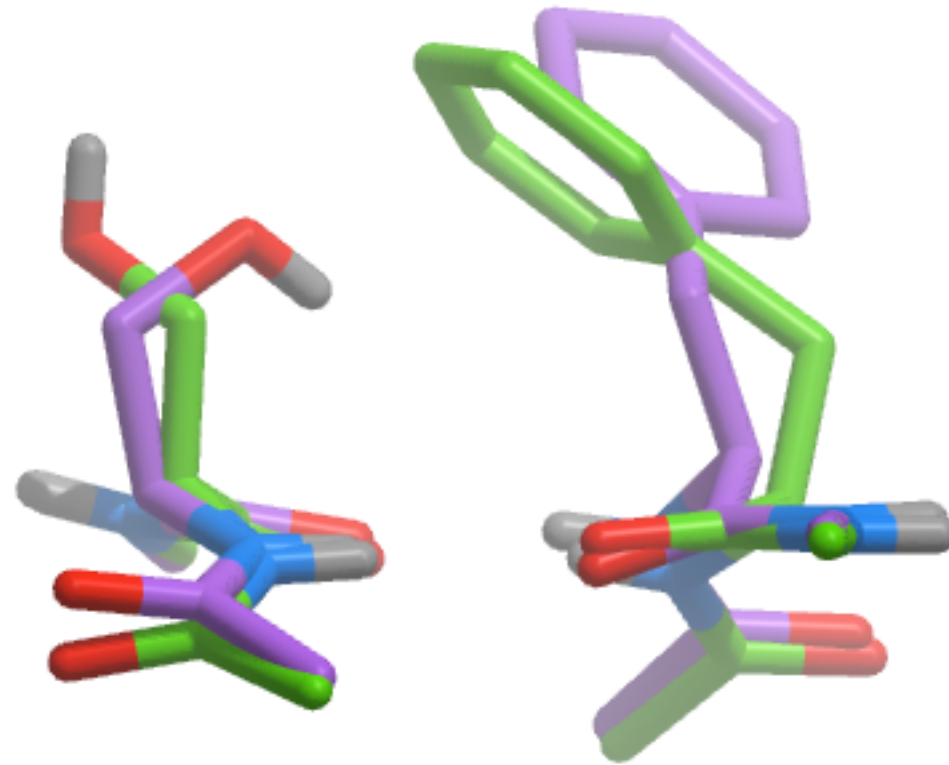
# Crystallography shows alternate conformations at room temperature



**Major Conformation (A)**

**Minor Conformation (B)**

# Crystallography shows alternate conformations at room temperature

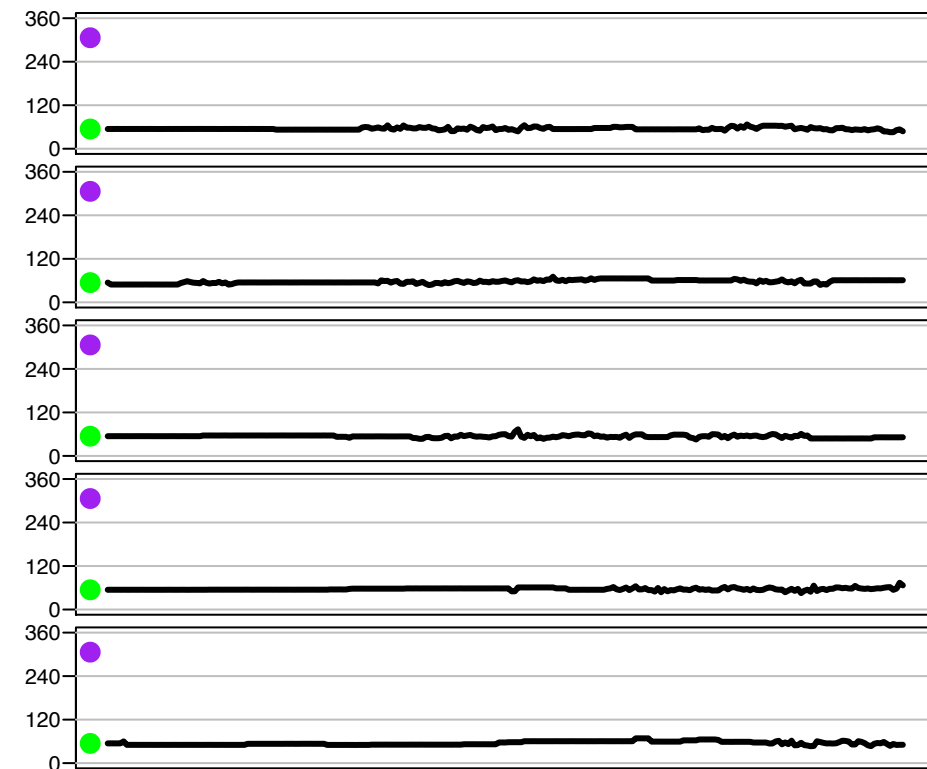
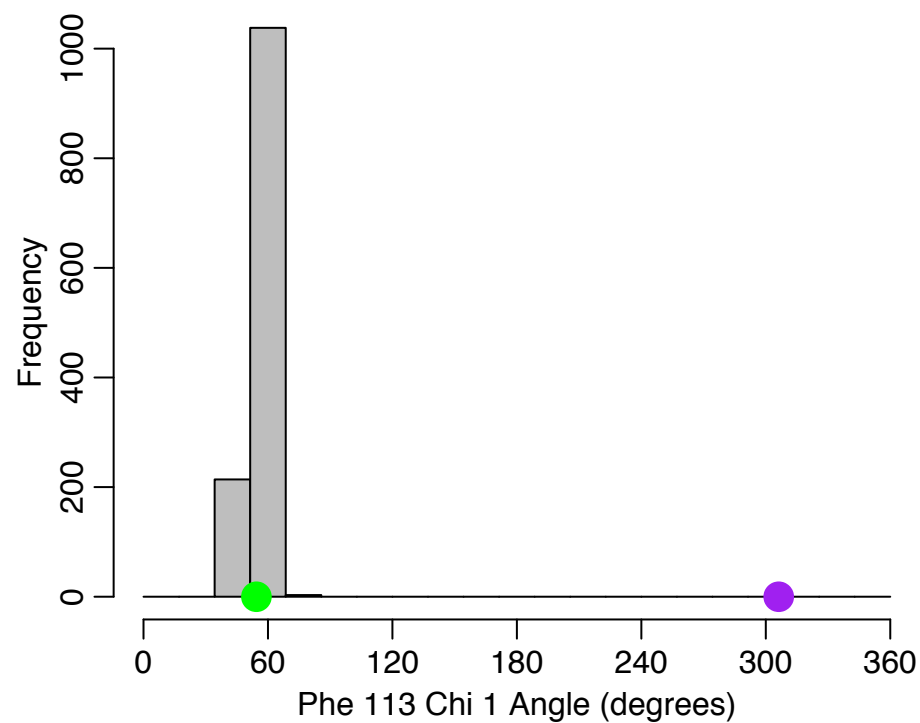
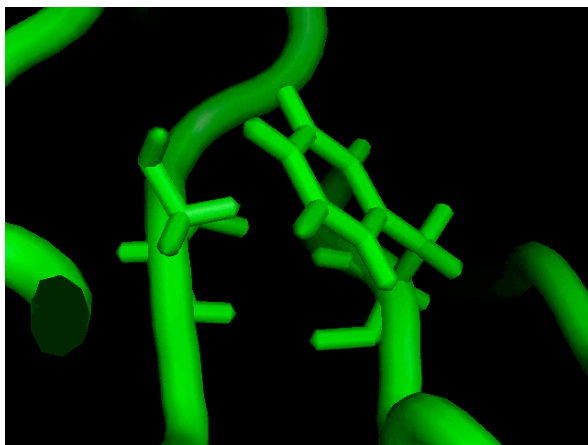


**Major Conformation (A)**

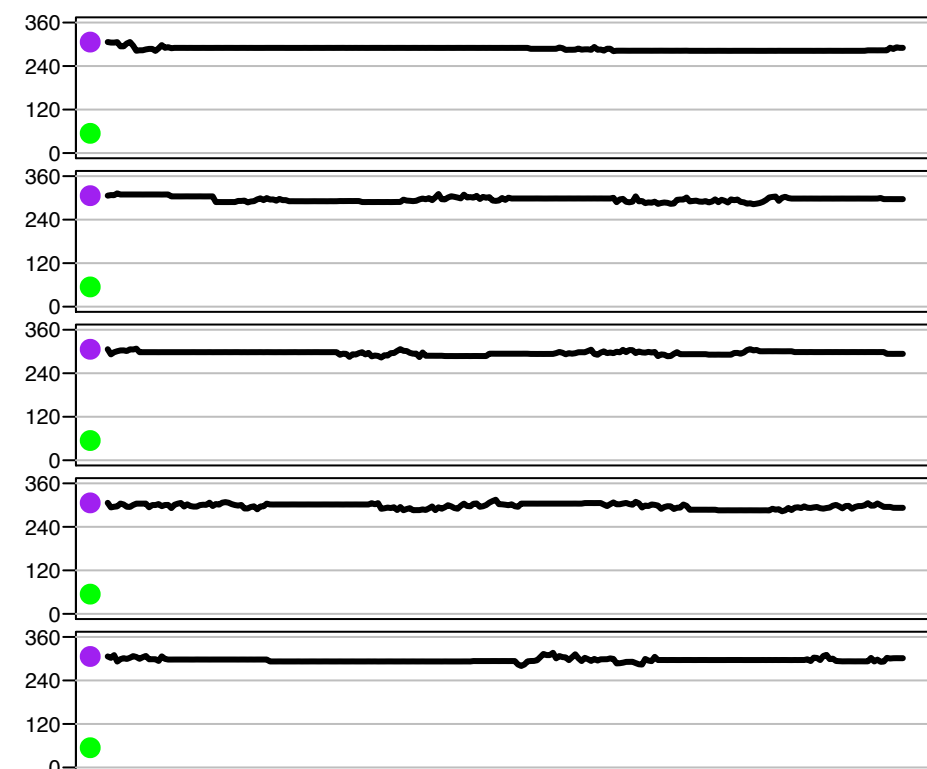
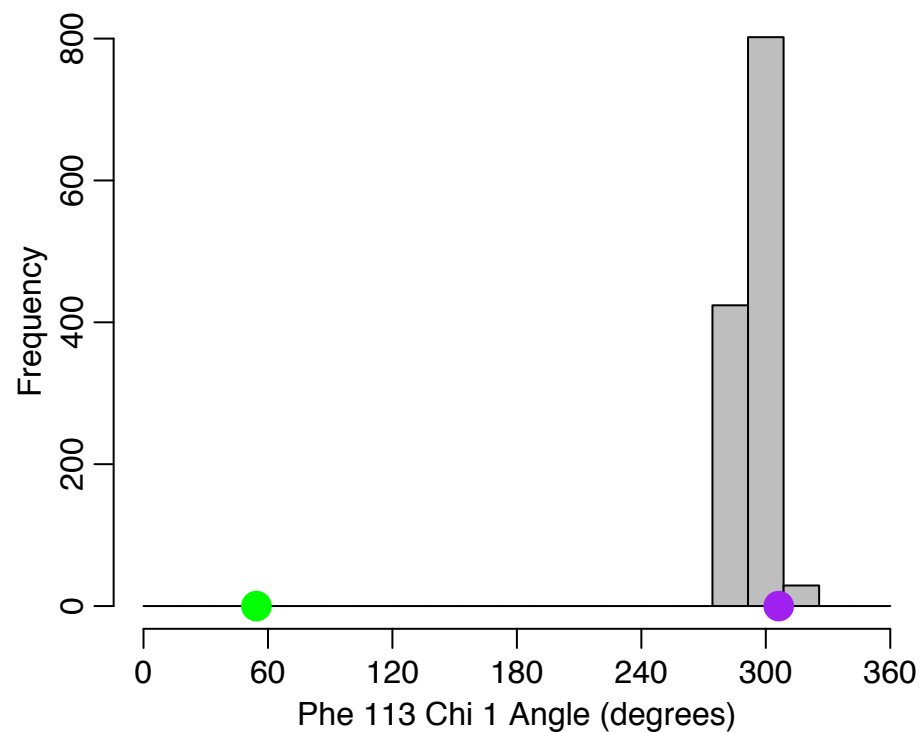
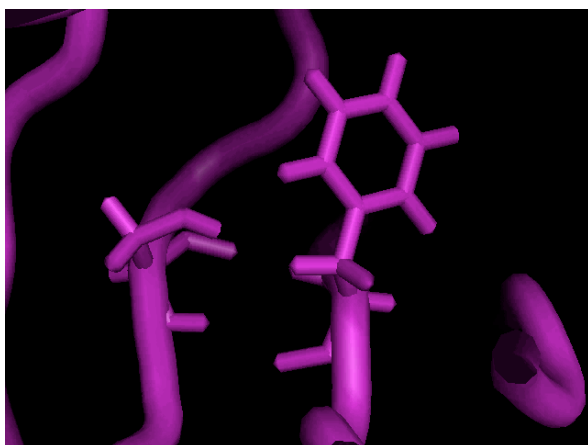
**Minor Conformation (B)**

# Little Phenylalanine Motion at 0.3 kT

Major (A)



Minor (B)

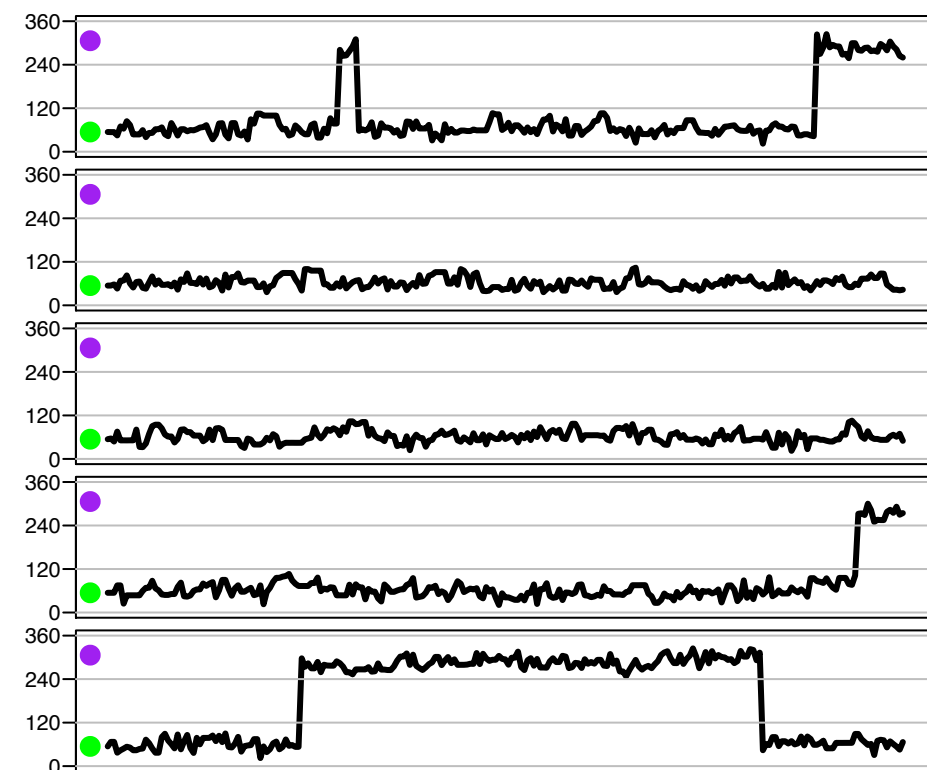
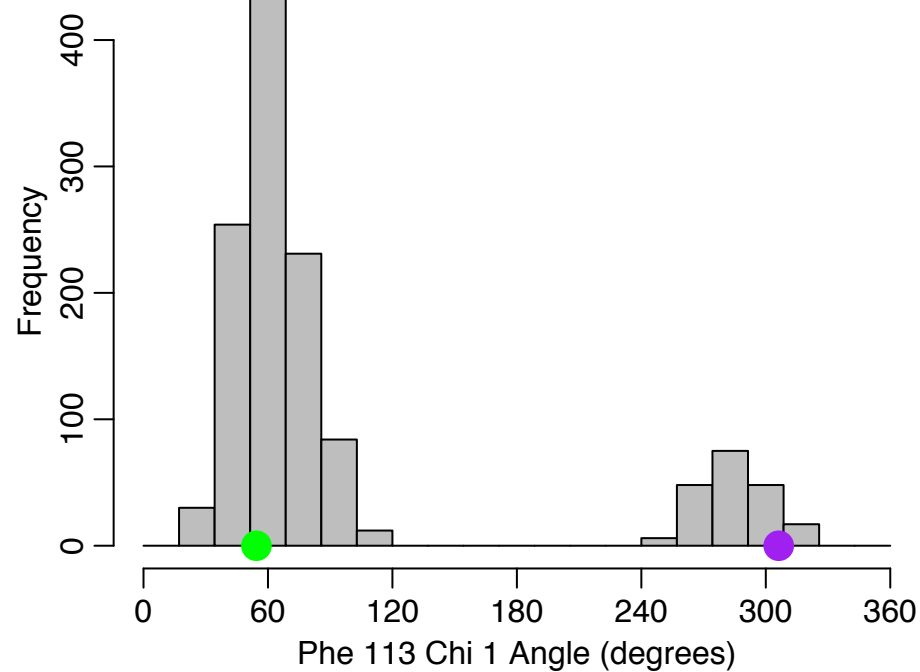
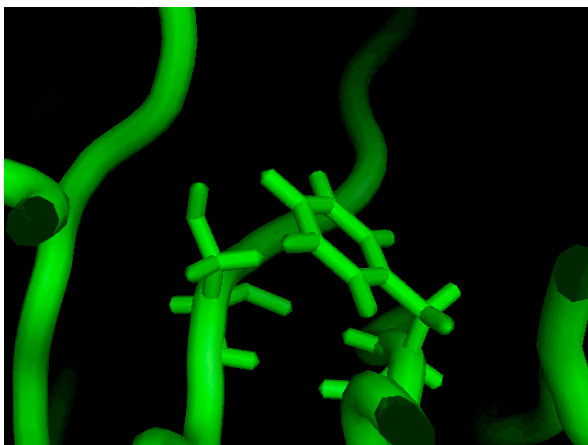


# Cyclophilin Protocol Changes

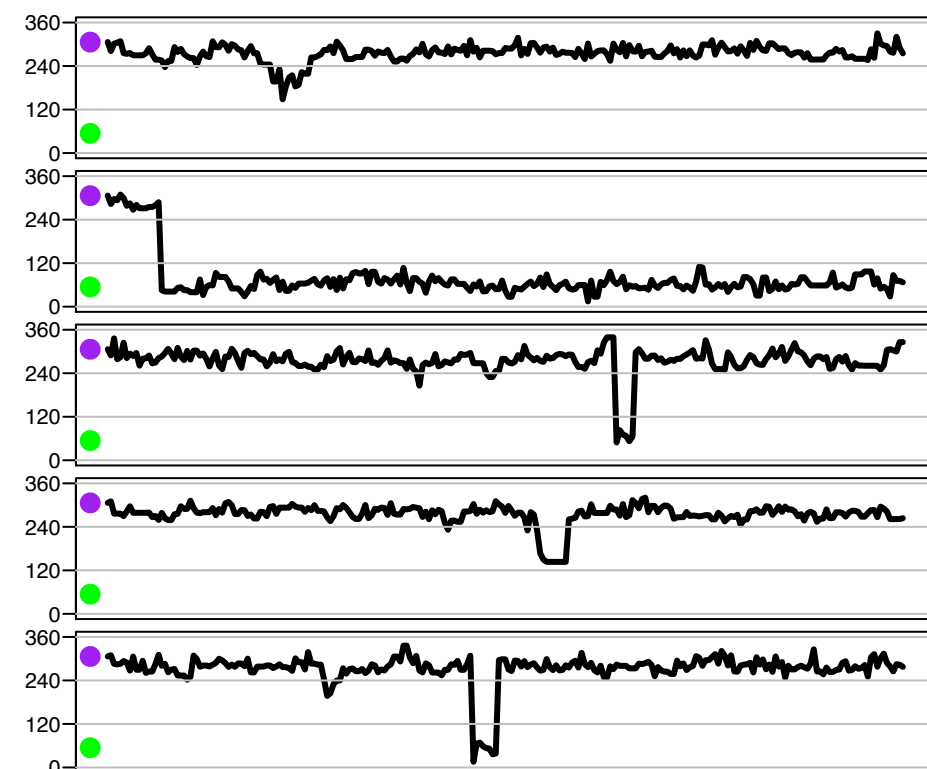
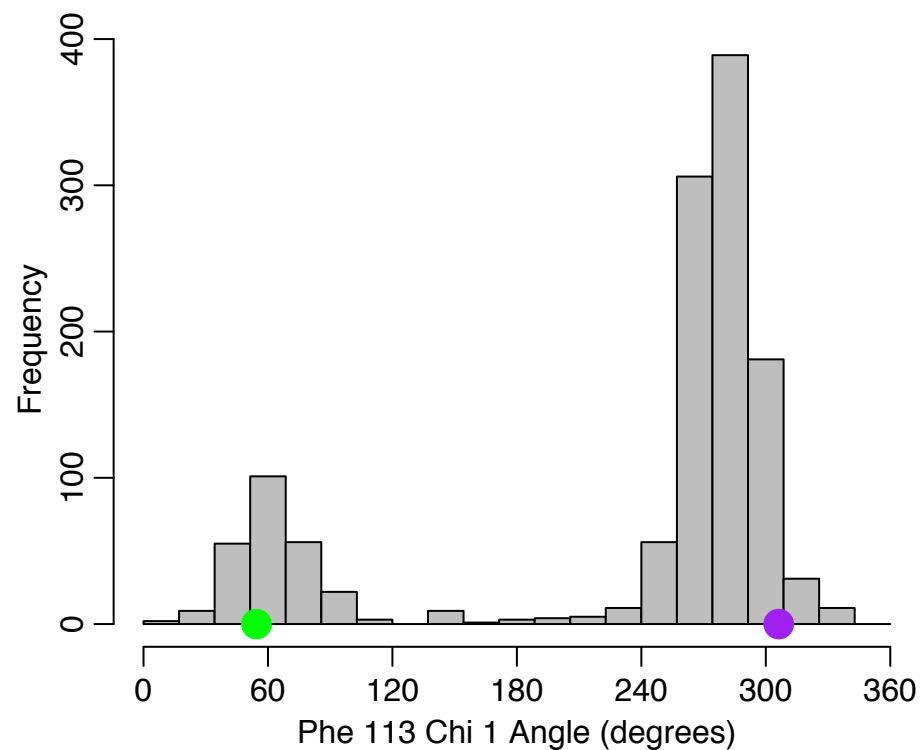
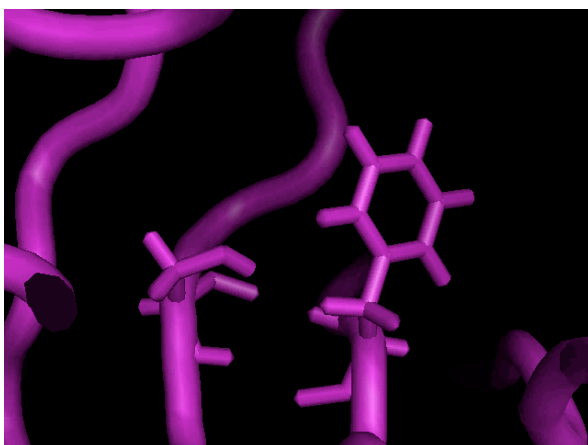
- Restricted sampling to 8 angstroms around Ser 99 & Phe 113
- Raised temperature to 4.8 kT (tried 0.6-2.4 along the way)
- Added simultaneous backrub side chain sampling
- Increased inter-rotamer sampling from 45% to 70%

# Phenylalanine Intraconverts at 4.8 kT

Major (A)



Minor (B)



# Thanks

Thomas Bliska (Williams College)

Tanja Kortemme (UCSF)

Jerome Nilmeier (UCB)

Yuan Liu (UW)

Liz Kellogg (UW)

Andrew Leaver-Fay (UNC)

David Baker (UW)

DOD

Genentech